RESEARCH ON THE FLAVONES CONTENT IN FOUR SPECIES OF MEDICINAL PLANTS GROWN IN THE WESTERN TRANSYLVANIAN PLAIN

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Abstract. Extracts of four species of Lamiaceae family medicinal plants (Agastache Foeniculum, Nepeta cataria, Melissa officinalis and Lavandula angustifolia) used in phytotherapy were screened for flavone content in two different harvest periods and two different hours of the day. Through this research we want to emphasize the optimal harvest time of vegetal raw material, when the studied plants contain the maximum active principles. The content of flavone/flavonols, main phenolics contained by these plants, was determined spectrophotometrically, according to Dowd method in APHIS Laboratory from USAMV Cluj-Napoca, with a Synergy 2 spectrophotometer, microplate reader. Duplicate samples of each plant were harvested for determinations from raw plant (fresh) and dry material. Following the data from Agastache Foeniculum and Nepeta cataria, the highest flavone content was obtained from the raw material harvested on 09/06/2014, at 16 hour, in both fresh and dried raw material. In the Melissa officinalis species, the highest flavone content was found in raw material (fresh and dry) which was harvested on 09/06/14, 11 hour. The highest flavone content in Lavandula angustifolia was found on 19/06/2014, 11 hour, in the fresh material and at 16 hour in the dry one. For the four studied species, the obtained results regarding the flavone content show that the optimal time of harvest was 09/06/2014, ie at the beginning of flowering, except for the species Melissa officinalis for which the flowering occured after 20/06/2014.

Keywords: Agastache foeniculum, Nepeta cataria, Melissa officinalis, Lavandula angustifolia, polyphenols.

INTRODUCTION

Dietary antioxidants have a great importance for the prevention of different diseases or health improvements and herbal preparations are a rich source of these compounds (Blasa et al., 2010; Bobiş et al., 2014; Turkmen et al., 2005).

Plant extracts obtained with different techniques and solvents are mixtures of several classes of compounds, rich in biologically active principles, represented by secondary metabolites synthesized in their growing process. The main bioactive compounds in plants are phenolic acids, free and glycosilated flavonoids (flavones, flavonols, dihydroflavones, flavan-3-ols etc.).

Lamiaceae family provide more than 200 plants genera. Special interest have Lavandula angustifolia, Melissa officinalis, Nepeta cataria and Agastache foeniculum for their high content of bioactive compounds.

Lavandula angustifolia is the only member of Lavandula genus which is recognized officially as a medicinal plant and used in phytotherapy (BLAŽEKOVIĆ et al., 2010). The chemical composition of this plant have been subjected to several studies over the past years, studies focused on the organic extracts, fractions of these extracts or essential oils extracted

from aerial parts or flowers of the plant (Blažeković et al., 2010; Hajhashemi et al., 2003; Hohmann et al., 1999; Lis-Balchin and Hart, 1999).

Melissa officinalis, popularly known as lemon balm is a medicinal plant having different properties in traditional medicine (treatment of gastrointestinal disturbance, minor sleeplessness, antioxidant and neuroprotective effects)(HANGANU ET AL., 2008; DUDA ET AL., 2007; ONIGA et al., 2010).

Nepeta cataria (catnip) leaves possess pleasant minty and lemony scent and taste and has been used in traditional medicine since long time. Also catnip is used for flavouring sauces, soups and cooked foods. It is used in phytomedicine as antispasmodic, antitussive, sedative, carminative, stomachic and tonic (DUKE, 1987; MIHAYLOVA et al., 2013). Nepeta cataria var. citriodora is mentioned as a possible adulteration or substitute for lemon balm (Melissa officinalis)(MODNICKI et al., 2007).

Agastache foeniculum is a herbaceous species, perennial, used in ornamental, medicinal and aromatic purposes (DUDA et al., 2013), known under the name "gyant hyssop". Among all medicinal plants, *A.foeniculum* presents an increasing interest in traditional phytotherapeutics, due to the essential oils composition and also non-volatile metabolites (ZIELINSKA AND MATKOWCKI, 2014).

The main objective of the present paper is to present the flavone content in the aerial parts from four species of *Lamiaceae* family (medicinal and aromatic plants) harvested in four different periods grown in the experimental field of UASVM Cluj-Napoca from Jucu.

AIMS

Through this research we wanted to emphasize the optimal harvest time of vegetal raw material, moment in which the maximum level of active principles are produced.

MATERIALS AND METHODS

The flavone/flavonol content of ethanolic extracts from *Agastache foeniculum, Nepeta cataria, Melissa officinalis* and *Lavandula angustifolia* was determined. These species of perennial medicinal plants were cultivated for approx. 7 years in the experimental field from Jucu of our university.

Plant samples were harvested in four steps, during two phenological periods: 9 and 19 of June 2014, in two different times of the day (11.00 and 16.00 hours).

Duplicate samples of each four species were collected - one for determination of active principles from fresh material, and the second sample was dried protected from light, and then analyzed.

Active principles determination from ethanolic extracts (2% in ethanol 70%) was carried out in APHIS Laboratory of USAMV Cluj-Napoca, using a Synergy 2 spectrophotometer, micro plate reader. Flavone/flavonol determination was performed following the Dowd method (Dowd, 1959), with some modifications.

A solution of aluminum chloride (2% in methanol) is used as a specific reagent for flavone/flavonol determination. In the presence of flavonoids the reagent develops a yellow-orange colour, proportionally with the amount of the compounds. The intensity of this color can be determined spectrophotometrically at 415 nm.

Work technique: a dilute ethanolic solution of each sample (100 μ L) was mixed with the same amount (100 μ L) of aluminum chloride 2% in methanol. Absorbance was recorded after 10 minutes of reaction. For flavone/flavonols quantification, a calibration curve of

different dilutions of quercetin $(0.005-0.1\ mg/mL)$ was performed $(y=7.435x+0.049,\ r^2=0.997)$.

Interpretation of the results for active principles (flavones) was performed by analysis of variance, Student's t test and Duncan test and that the two-factor statistical calculation program « ANOVA» of the discipline Experimental Techniques in our university.

RESULTS

Flavones and flavonols are plant derived polyphenolic compounds useful in human diet. They are presented in fruits, vegetables, tea, red wine and medicinal and aromatic herbs. Most flavonoids are known to be chemo-preventive, lowering risks of cancer and other diseases. Medicinal plants from *Lamiaceae* family are known to have high amounts of polyphenols in general and consequently flavonoids also. The main problem for harvesting these benefic plants is finding the optimum moment, when these compounds have the highest accumulation in different parts of the plant. The present study tried to find this optimal moment, by harvesting four species of *Lamiaceae* family in two different phonological stages, and in two different hours of the day. Results show that these four periods show different amounts of flavones, and give the answer regarding the optimal moment of harvest, in order to have the highest amount of active principle.

Tables 1 and 2 present the results of flavone content from *Agastache foeniculum* (extract made from fresh and dry plant).

Table 1 Flavone content of fresh aerial parts from Agastache foeniculum

Date	Time of	Concentration	Diff	erences	Significance	Duncan
	harvest	mg/ml	%	Concentrati on mg/ml		Test
09/06/2014	11	9.42	100.0	0.00	Control	A
	16	10.99	116.8	1.58	***	С
19/06/2014	11	10.95	116.3	1.53	***	С
	16	10.22	108.5	0.80	***	В
DL(p 5%) = 0.35; DL(p 1%) = 0.48; DL(p 0.1%) = 0.68						

Table 2

Table 3

Flavone content of the dried aerial parts from Agastache foeniculum

Date	Time of	Concentration	D	ifferences	C::f:	Duncan	
	harvest	st mg/ml	%	Concentration mg/ml	Significance	Test	
00/06/2014	11	16.40	100.0	0.00	Control	В	
09/06/2014	16	19.06	116.2	2.66	***	С	
19/06/2014	11	15.89	96.9	-0.52	00	A	
	16	16.30	99.4	-0.10	-	В	
DL(p 5%) = 0.35; $DL(p 1%) = 0.48;$ $DL(p 0.1%) = 0.68$							

Obtained results were in accordance with literature study findings, even if the extraction methods and the way of results expression were sometimes different (DAPKEVICIUS ET AL., 1998; ZIELINSKA AND MATKOWCKI, 2014).

Positive very significant differences regarding the content of flavones from dry material, compared to the control (sample harvested 09/06/2014, 11 hours) was obtained from raw materials harvested on 09/06/2014, 16 hours.

Interpreting the same results also through the multiple comparisons method (Duncan test) it can be observed that the highest content of flavanols is obtained from raw materials harvested in early flowering, at 16 hours.

The content of flavones from Nepeta cataria (catnip) is presented in tables 3 and 4.

Through the analysis of variance is established a very significant difference in fresh material and distinct significant in dry material compared to the control (09/06/2014, 16 hours) regarding the content of flavones.

By interpreting the data using the Duncan test we found that the best time to harvest is the beginning of flowering (for the last year, 9th of June), in the afternoon (16 hours), both for fresh and dry material.

Flavone content of fresh aerial parts from *Nepeta cataria* (catnip)

Pravone content of fresh acrial parts from wepen cultura (camp)								
Date	Time of	Concentration	D	ifferences	C::f:	Duncan		
	harvest	mg/ml	%	Concentration mg/ml	Significance	Test		
09/06/2014	11	2.64	100.0	0.00	Control	С		
	16	2.99	113.2	0.35	***	D		
19/06/2014	11	1.80	68.2	-0.84	000	A		
	16	1.89	71.3	-0.76	000	В		

DL(p 5%) = 0.07; DL(p 1%) = 0.10; DL(p 0.1%) = 0.16

Table 4 Flavone content of the dried aerial parts from Nepeta cataria (catnip)

Date	Time of Concentration		Differences		ac.	Duncan
	harvest	mg/ml	%	Concentration mg/ml	Significance	Test
00/06/2014	11	8.07	100.0	0.00	Control	В
09/06/2014	16	8.48	105.0	0.40	**	С
19/06/2014	11	5.91	73.2	-2.17	000	A
	16	6.05	74.9	-2.03	000	A
DL(p 5%) = 0.18; $DL(p 1%) = 0.27;$ $DL(p 0.1%) = 0.44$						

Centralization and interpretation of statistical data on the content of flavones from *Melissa officinalis* (lemon balm) is shown in Tables 5 and 6.

Through the analysis of variance is established a significant very negative difference at all moments of harvest compared to the control (09/06/2014, 11 hours) for fresh raw material. In terms of dry sample material differences are significantly negative for the samples collected on 9 and 19/06/2014, 16 hours, compared to control (09/06/2014, 11 hours).

The highest content of flavones is in the raw material (fresh and dry) harvested at the beginning of flowering (09/06/2014), 11 hours.

Table 5
Flavone content of fresh aerial parts from *Melissa officinalis* (lemon halm)

Fla	Flavone content of fresh aerial parts from <i>Metissa officinatis</i> (lemon balin)								
Date	Time of	Concentration		oifferences	C::C:	Duncan			
	harvest	mg/ml	%	Concentration mg/ml	Significance	Test			
	11	2.94	100.0	0.00	Control				
09/06/2014	11					D			
	16	2.40	81.5	-0.54	000	В			
19/06/2014	11	2.56	87.2	-0.38	000	С			
	16	2.23	75.9	-0.71	000	A			
DI (n	DI(p.5%) = 0.07; $DI(p.1%) = 0.11$; $DI(p.0.1%) = 0.18$								

Table 6
Flavone content of the dried aerial parts from *Melissa officinalis* (lemon balm)

	Flavone content of the dried aerial parts from <i>Metissa officinatis</i> (lefifon ballit)								
Date	Time	Concentration	Г	Differences	C::C:	Duncan			
	of harvest	mg/ml	%	Concentration mg/ml	Significance	Test			
00/06/2014	11	5.93	100.0	0.00	Control	В			
09/06/2014	16	5.49	92.5	-0.44	0	A			
19/06/2014	11	5.60	94.5	-0.33	-	AB			
	16	5.55	93.6	-0.38	0	A			
DL (p 5%) = 0.34; DL (p 1%) = 0.51; DL (p 0.1%) = 0.82									

In Tables 7 and 8 are centralized and interpreted statistical data on the content of flavones from Lavandula angustifolia.

It can be observed very low concentrations of flavones, with significant negative differences compared to the control (09/06/2014, 11 hours) to fresh aerial parts. In contrast to fresh feedstock, at the dry matter content it is noticed a very high concentration in flavones on 09/06/2014, 16 hours, with positive significant differences compared to the control.

By statistialy interpreting the data using the Duncan test it appears that the optimal time for harvesting samples is at early flowering (last year was 9th of June), before noon, at 11 hour, in the case of fresh ones and in the afternoon, at 16 hours in the case of raw dried material regarding the content of flavones.

Table 7 Flavone content of fresh aerial parts from Lavandula angustifolia (lavender)

Date	Time of	Concentration	D	ifferences	aa.	Duncan	
		mg/ml	%	Concentration mg/ml	Significance	Test	
	11	1.75	100.0	0.00	Control		
06/09/2014	11					D	
	16	1.62	92.6	-0.13	000	В	
06/19/2014	11	1.66	94.7	-0.09	000	С	
	16	1.39	79.7	-0.35	000	A	
DL(p 5%) = 0.03; DL(p 1%) = 0.05; DL(p 0.1%) = 0.08							

Table 8 Flavone content of the dried aerial parts from Lavandula angustifolia (lavender)

Thavone content of the dried defini parts from Edvandana angustyona (lavender)							
Date	Time of	Concentration	D	ifferences	C::f:	Duncan	
	harvest	mg/ml	%	Concentration mg/ml	Significance	Test	
0.1/0.0/2.01	11	3.11	100.0	0.00	Control	В	
06/09/2014	16	3.61	116.1	0.50	***	D	
06/19/2014	11	3.39	109.2	0.28	**	С	
	16	2.90	93.2	-0.21	0	A	
DL (p 5%) = 0.16; DL (p 1%) = 0.24; DL (p 0.1%) = 0.39							

The concentrations of flavones for the 4 species in the four moments of sampling set according to the research protocol, such as the production of fresh and dry matter, is shown in Figures 1 and 2.

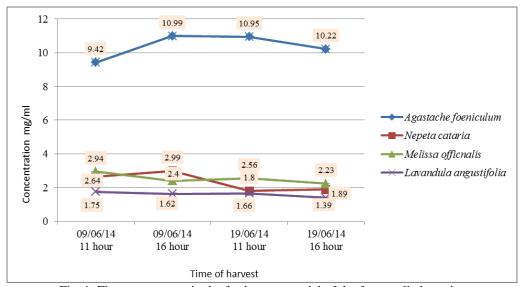


Fig. 1. Flavones content in the fresh raw material of the four studied species

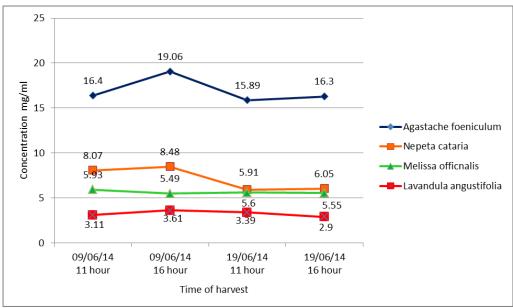


Fig. 2. Flavones content in dry matter raw material of the four studied species

Following the data from *Agastache foeniculum* and *Nepeta cataria* we can conclude that the highest flavone content was obtained from the raw material harvested on 09/06/2014, at 16 hours, in both fresh and dried raw material. In the *Melissa officinalis* species, the highest flavone content was found in the raw material (fresh and dry) which was harvested about 10 days before blooming on 09/06/14, 11 hour. The highest flavone content in *Lavandula angustifolia* was found at the starting period of blooming, in the case of 2014 on the 9th of June, 11 hour, for the fresh material and at 16 hour for the dry one.

CONCLUSION

The optimal harvest time, when the flavones concentration was highest, was found to be the beginning of flowering (09/06/2014), at 16 hour in the afternoon, in three from the four species studied, ie *Agastache foeniculum*, *Nepeta cataria* and *Lavandula angustifolia*, except *Melissa officinalis* species, for which the maximum concentration in flavones was the same date (09/06/2014), earlier in the morning, at 11 hour. On this date lemon balm was approx. 10 days before the starting of flowering.

Of the four species analyzed, the highest concentrations in flavones were recorded at *Agastache Foeniculum*, of 3-5 times higher than in other species, both at fresh and dry material.

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BIBLIOGRAPHY

- 1. BLASA, M., GENNARY, L., ANGELINO, D., NINGALI, P., 2010, Fruit and vegetable antioxidants in health. In: Watson, R., Preedy, V. (Eds.). Bioactive Foods in Promoting Health Fruits and Vegetables. Academic Press, Massachusetts. p.37-58.
- 2. MIHAYLOVA, D., GEORGIEVA, L., PAVLOV, A., 2013, In vitro Antioxidant activity and phenolic composition of *Nepeta cataria* L., extracts, International Journal of Agricultural Science and Technology, 1(4), 74-79.
- 3. DUKE, L.A., 1085, Handbook of medicinal herbs, 2ed, CRC Press, Boca Raton, Florida, USA.
- 4. BLAŽEKOVIĆ, B., VLADIMIR-KNEŽEVIĆ, S., BRANTNER, A., BIVAL ŠTEFAN, M., 2010, Evaluation of antioxidant potential of *Lavandula x intermedia* Emeric ex. Loise. `Budrovka`: a comparative study with *L. angustifolia* Mill., *Molecules*, 15, 5971-5987.
- 5. BOBIŞ, O., DEZMIREAN, D.S., TOMOŞ, L., CHIRILĂ, F, MĂRGHITAŞ, L.A., 2015, Influence of phytochemical profile on antibacterial activity of different medicinal plants against gram-positive and gram-negative bacteria, *Applied Biochemistry and Microbiology*, 51(1), 113-118.
- 6. DOWD, L.E., 1959, Spectrophotometric determination of quercetin, *Analytical Chemistry*, 31(7), 1184-1187
- 7. DUDA, M., VÂRBAN, D., CĂTANĂ, C., MUNTEAN, S., BANGA, D., 2007, Ways to extract free compounds from *Melissa officinalis* L., 1st International Scientific Conference on Medicinal Aromatic and Spice Plants, Book of Scientific Papers, Nitra, 119-121.
- 8. HAJHASHEMI, V., GHANNADI, A., SHARIF, B., 2003, Anti-inflammatory and analgesic properties of the leaf extracts and essential oil of *Lavandula angustifolia* Mill., Journal of Ethnopharmacology, 89, 67-71.
- 9. HANGANU, D., VLASE, L., FILIP, L., SAND, C., MIREL, S., INDREI, L., 2008, The study of some polyphenolic compounds from *Melissa officinalis* L. (Lamiaceae), *Revista Medico-Chirurgicală a Societății de Medici și Naturaliști din Iași*, 2(2), 523-527.
- 10. HOHMANN, J., ZUPKO, I, REDEI, D., CSANY, M., FALKAY, G., MATHE, I., 1999, Protective effects of the aerial parts of *Salvia officinalis, Melissa officinalis* and *Lavandula angustif*olia and their constituents against enzyme-dependent and enzyme-independent lipid peroxidation. *Planta Medica*, 65, 576-578.
- 11. LIS-BALCHIN, M., HART, S., 1999, Studies on the mode of action of the essential oil lavender (*Lavandula angustifolia* P. Miller), *Phytotherapy Research*, 13, 540-542.
- 12. MUNTEAN, L.S., M. TĂMAŞ, S. MUNTEAN, L. MUNTEAN, M.M. DUDA, D.I. VÂRBAN, S. FLORIAN, 2007. Tratat de plante medicinale cultivate şi spontane, Ed. Risoprint Cluj-Napoca, 928 p, ISBN 978-973-751-463-9.
- 13. ONIGA, I., VLASE, L., TOIU, A., BENEDEC, D., DUDA, M., 2010, Evaluation of phenolic acid derivatives and essential oil content in some *Melissa officinalis* L. varieties, *Farmacia*, 58(6), 764-769.
- 14. TURKMEN, N., SARI, F., VELIOGLU, Y.S., 2005, The effect of cooking methods on total phenolics and antioxidant activity of selected green vegetables, *Food Chemistry*, 93, 713-718.
- 15. MODNICKI, D., TOKAR, M., KLIMEK, B., 2007, Flavonoids and phenolic acids of *Nepeta cataria* L. var. citriodora (Becker) Balb. (*Lamiaceae*), *Acta Poloniae Pharmaceutica Drug Research*, 64(3), 247-252.

- 16. DUDA, M.M., MATEI, C.F., VÂRBAN, S.M., MOLDOVAN, C., 2013, The results of cultivating the species *Agastache foeniculum* (Pursh) Kuntze at Jucu, CJ, *Bulletin USAMV serie Agriculture*, 70(1):214-217.
- 17. ZIELINSKA, S., MATKOWSKI, A., 2014, Phytochemistry and bioactivity of aromatic and medicinal plants from the genus *Agastache* (Lamiaceae), Phytochemical Review, 13, 391-416.
- 18. DAPKEVICIUS, A., VENSKUTONIS, R., VAN BEEK, T.A., 1998, Antioxidant activity of extracts obtained by different isolation procedures from some aromatic herbs grown in Lithuania, *Journal of the Science of Food and Agriculture*, 77, 140-146.