INFLUENCE OF CROP MANAGEMENT ON THE IMPACT OF ZYMOSEPTORIA TRITICI IN WINTER WHEAT IN THE CONTEXT OF CLIMATE CHANGE: AN OVERVIEW

Otilia COTUNA¹, Mirela PARASCHIVU², M. PARASCHIVU², L. OLARU²

¹Banat`s University of Agricultural Sciences and Veterinary Medicine "King Michael I of Romania"
Timisoara, Romania

²University of Craiova, Agronomy Faculty, Craiova, Romania

Corresponding author: otiliacotuna@yahoo.com; paraschivumirela@yahoo.com paraschivum@yahoo.com; liviu.olarudtas@gmail.com

Abstract. The impact of climate change on specific biotic constrainers like pathogens and on the host-pathogen relationship is associated with changes in pathogens life cycles, increased incidence, pathogenicity, genetically recombination and aggressiveness traits, which require rethinking the integrated management strategies. However, the results of investigations are inconsistent and poorly understood in the context of climatic change. The present review is focused on the influence of crop management on Septoria Tritici Blotch (STB) disease in winter wheat in the context of climate change taking into consideration case studies in order to understand better how the components of disease cycle are affected and to identify disease risk of different agricultural practices. The response of Zymoseptoria tritici pathogen to climate change is of high interest currently in order to estimate disease risk on a large scale and to introduce new understandings in developing management strategies. Soil tillage, crop rotation, sowing date and nitrogen fertilization are considered important tools in disease control which need to be adjusted according with climatically factors for each area which affect the dispersal of ascospores and pycnidiospores of the pathogen. However, further investigation need to be done in order to highlight the impact of climate change on foliar wheat pathogens and which are the most appropriate management tools in order to control these pathogens and to enhance global food security in a changing climate.

Key words: Septoria Tritici Blotch (STB), climate change, crop management, host-pathogen interactions, Zymoseptoria tritici.

INTRODUCTION

The intensification of world agriculture has to happen in a time, when climate is becoming less predictable, fossil fuel dependency needs to be cut, and cropland and water resources are deteriorating [ALEXANDRATOS AND BRUINSMA, 2012; POPP et al., 2013]. There are scenarios that climatic change will lead to a higher incidence of crop diseases (especially plant host and susceptibility, pathogen reproduction – shorter incubation - dispersal, survival and activity, host-pathogen relationship) and to a potentially larger use of pesticides [NEWTON et al., 2011; SUTHERST et al., 2011]. The predicted impact of climate change on pathogens and host-pathogen relationship suggests that can be positive, negative or neutral depending on geographical and temporal distribution of inoculum amount and cultivars susceptibility [NEWTON et al., 2011; SUTHERST et al., 2011]. Thus, new pathogens may occur in certain regions, while other pathogens may decreases to be economically important, following geographical distribution of the host and cropping technology [COAKLEY et al., 1999; GHINI et

al., 2008; GHINI AND HAMADA, 2008]. General tendency is that pathogens are likely to remain limited to their host distribution and not become disconnected from them. There is serious concern that climate zones will move faster that it is possible for plant populations to track them, which is expected to determine disproportionate extinction of local endemic species [LOARIE et al., 2009]. However, advanced knowledge about the impact of climate change on plant diseases is essential for adoption of integrated disease management measures, in order to avoid yield losses.

The ascomycete *Zymoseptoria tritici* (Desm.) Quaedvlieg & Crous (synonymous *Septoria tritici*; telemorph *Mycosphaerella graminicola*) is known as one of the most devastating foliar diseases of wheat worldwide in different climatic areas, causing *Septoria Tritici* Blotch (STB) disease [DEAN *et al.*, 2012; JØRGENSEN *et al.*, 2014; GURR AND FONES, 2015]. Also, *Zymoseptoria tritici* has been recorded on 26 grasses and non-graminaceous weed, but among all these only six grass species have been validated as alternative hosts of *Zymoseptoria tritici* [PRESTES AND HENDRIX, 1978; SUFFERT *et al.*, 2011].

Most studies on *Zymoseptoria tritici* are mainly focused on the host-pathogen interaction and pathogen management, but recently many authors pay a special attention to the pathogen cell biology, like cellular organization and intercellular dynamics of the pathogen itself, [Mehrabi *et al.*, 2006; Orton *et al.*, 2011; Suffert *et al.*, 2011; Steinberg G., 2015] and fungus genetics [Mehrabi *et al.*, 2006, 2009; Stukenbrock *et al.*, 2006, 2010; Wittenberg *et al.*, 2009; Godwin *et al.*, 2011; Dhillon *et al.*, 2014; Kellner *et al.* 2014; Mirzadi Gohari *et al.*, 2014; Kettles and Kanyuka, 2016; Saintenac *et al.*, 2018].

Several studies have been made on host-pathogen interaction in the context of climate change [GLADDERS et al., 2001; PIETRAVALLE et al., 2003], but despite the progress in genetic resistance and fungicide control the pathogen has proven adapt to overcoming resistance and develop fungicide resistance, therefore new approaches are needed to be integrated in disease management. Currently, there is no full genetic resistance in wheat to *Zymoseptoria tritici*, so *Septoria Tritici* Blotch (STB) disease is mainly managed by fungicides most of the cases, being estimated that 70% of the total annual fungicide used in European Union is used for this pathogen [PONOMARENKO et al., 2011; TORRIANI et al., 2015]. There was estimated that STB disease could cost the economies of France, Germany and the UK, the three main EU wheat growing countries, between €120 and 700 million [GURR AND FONES, 2015]. Despite that general adopted levels of yield losses worldwide (up to 50% of yield during severe epidemics) [EYAL et al., 1973, 1987] new information are necessary to be considered on specific areas under the impact of climatic changes. However, *Zymoseptoria tritici* is a pathogen that can lead to an epidemic depending on the prevalent environmental variables. The impact of climate change on the fungus *Zymoseptoria tritici* and its management is relatively understudied.

This paper aims to give an overview of the current knowledge of the impact of crop management on the fungus *Zymoseptoria tritici* in the context of climate change, which makes wheat yield more vulnerable and threaten food security. Understanding the relationship between environment conditions, crop management practices and spore dispersal could improve the control strategy of *Septoria Tritici* Blotch (STB) disease.

CLIMATICAL FACTORS INFLUENCE ON $ZYMOSEPTORIA\ TRITICI$ IN WHEAT

The impact of climate change needs to be considered along with other factors that affect crop yields, such as specific biotic constrainers (pathogens) and its impact on the host-pathogen relationship.

Warmer temperatures are expected to occur in the north regions leading to extended areas cropped with cereals which will increase the diseases incidence and severity [GHINI et al., 2008]. There were formulated three hypotheses how pathogens will be influenced by rising temperatures under climate change scenario: (i) an increased pathogen development transmission and generation number; (ii) an increased overwinter survival and reduced growth restrictions during this period and (iii) an alteration of host susceptibility [HARVELL et al., 2002]. GOUACHE et al., (2013) suggest that climatic change might reduce the average severity of STB by 2 - 6% in some areas in France, and to result in more low severity years and fewer high severity years (the probability that average severity will increase rather than decrease is between 18 - 45%, depending on location).

Among the 21 chromosomes carried by *Zymoseptoria tritici* eight of the smallest ones which are considered dispensable are supposed to be involved in faster adaptation of the pathogen to climatic changes [WITTENBERG et al., 2009; GOODWIN et al., 2011; SAINTENAC C. et al., 2018], fungicide resistance [TORRIANI et al., 2009; COOLS AND FRAAIJE, 2013], overcoming disease host resistance [MUNDT et al., 1999, MUNDT et al., 2002, RUDD et al., 2015].

Several studies have been made on the impact of environmental factors (radiation, temperature, rainfall, relative humidity, atmospheric carbon dioxide (CO₂) and other greenhouse gases) on Zymoseptoria tritici, suggesting that the most important role is played by temperature fluctuations, because leaves temperature is actually the temperature of the pathogen's body that develop onto or into plant leaves, influencing significantly their life cycle [GLADDERS et al., 2001; PIETRAVALLE et al., 2003; LOVELL et al., 2004; BEEST et al., 2009]. Warmer temperatures during early autumn will expose seedlings to an increased risk of infection with pathogen ascospores under sufficient moisture on leaves, especially on susceptible winter wheat cultivars, leading to a higher amount of inoculum overwinter. Thus, the survival of ascospores on unburied crop residues will play a large role in determining subsequent epidemic severity. Subsequent disease development results from pycnidiospores splashed vertically by rain infecting upper leaves close to each other completing several successive infections [SCHUN, 1990; LOVELL et al., 1997; CORDO et al., 1999; MCDONALD et al. 1999; SUFFERT et al., 2011]. Pycnidiospores are carried by asexual fruiting bodies called pycnidia, developed within the substomatal cavity on wheat leaf tissues which are covered with chlorotic and necrotic lesions [KEMA et al.,, 1996; DUNCAN AND HOWARD, 2000].

Beside temperature the moist leaf surface plays an important role for early infections being necessary an amount of rainfall of 10 mm three consecutive wet days with at least 1 mm rain [PIETRAVALLE *et al.*, 2003]. However, the pycnidiospores realise was associated with the increase in rainfall intensity seven days before the occurrence of the event and the increase in radiation 60 days before the same event. CORDO *et al.* (2017) showed that relative humidity three or five days before event was positively correlated with ascospores release and negatively correlated with radiation and temperature.

Regarding the effect of CO_2 on STB disease, VÁRY et al. (2015) showed that elevated CO_2 increased the severity of the disease and the acclimation of the pathogens leading to higher epidemics.

WHEAT CROP MNAGEMENT IMPACT ON ZYMOSEPTORIA TRITICI UNDER CLIMATE CHANGE

Despite contradictory results worldwide, agronomic factors are considered efficient tools to induce a break down into pathogens life cycle.

Beside altered weather patterns wheat crop practices will influence *Septoria Tritici* Blotch (STB) disease severity. Some studies emphasised that when effective management practices, including fungicide treatments, applied under no-till systems in wheat monoculture, the occurrence of *Septoria Tritici* Blotch (STB) is possible without significant yield losses [CARMONA *et al.*, 1999].

In general, among crop management practices, crop rotation is considered very efficient in decreasing wheat foliar diseases and fungicide pressure, especially in no-till systems, but in case of STB short crop rotation does not decrease significantly the disease level because *Zymoseptoria tritici* pycnidiospores are more important for disease spreading and development than primary inoculum existing on plant debris [BANKINA *et al.*, 2014; FERNANDEZ *et al.*, 2016; WENDA - PIESIK *et al.*, 2016].

No much work has been done on the effects of minimum tillage and no tillage on STB epidemics. The impact of soil tillage on *Zymoseptoria tritici* has been studied in few different geographical areas leading to contradictory results [BAILEY *et al.*, 2001; DUVIVIER *et al.*, 2013; BANKINA *et al.*, 2014]. The severity of *Septoria Tritici* Blotch (STB) disease was higher in ploughed plots under conventional tillage [GILBERT AND WOODS, 2001; BÜRGER *et al.*, 2012; FERNANDEZ *et al.*, 2016] then in other tillage systems.

Also, delaying autumn sowing date may reduce the severity of STB disease due to the fact that wheat crop may escape from ascospores primary infections [LOVELL *et al.*, 1997: GLADDERS *et al.*, 2001].

Nitrogen fertilization may affect STB disease by increasing wheat tissues susceptibility and by increasing plants canopy bringing closer wheat leaves one to each other which make it easier rain splash spores dispersal and by influencing the pathogen's life cycle itself [LOVELL *et al.*, 2007; WALTERS AND BINGHAMANN, 2007; PONOMARENKO *et al.*, 2011].

CONCLUSIONS

Wheat foliar pathogens are important yield-reducing components, but under climatic change conditions their attacks are likely to be more unpredictable and the amplitude larger. New studies on the impact of climate changes on foliar wheat diseases are required to make good predictions in order to design solutions to new problems challenges or to old problems becoming more severe.

Septoria Tritici Blotch (STB) disease, caused by Zymoseptoria tritici, is considered the most prevalent and devastating wheat disease in many countries worldwide. Studies of the impact of agronomic factors on the development of Septoria Tritici blotch (STB) disease have revealed high variations especially in the effect of crop rotation and soil tillage. However, some studies showed that the development of Septoria Tritici Blotch (STB) disease mainly

depends on climatic variables and less on agronomic practices, which make this disease more difficult to be managed under climate change conditions.

A better understanding of the interractions among climatic factors, pathogens biology and crop management of susceptible/resistant cultivars are keys to managing STB disease accross different geographical areas.

BIBLIOGRAPHY

- 1. ALEXANDRATOS, N., J. BRUINSMA, 2012, World agriculture towards 2030/2050: the 2012 revision. *ESA Working paper* No. 12-03. Rome, FAO;
- 2. BAILEY K. L., GOSSEN B. D., LAFOND G. P., WATSON, P. R., DERKSEN, D. A., 2001, Effect of tillage and crop rotation on root and foliar diseases of wheat and pea in Saskatchewan from 1001-1998: Unvariate and Multivariate analysis. *Can.J. Plat. Sci.* 81:789-803;
- 3. BANKINA, B., GAILE, Z., BALODIS, O., BIMŠTEINE, G., KATAMADZE, M., KREITA, D., PAURA, L., PRIEKULE, I., 2014 Harmful winter wheat diseases and possibilities for their integrated control in Latvia. *Acta Agric. Scand. Sect. B Soil Plant Sci.* 64:615–622;
- 4. BÜRGER, J., GÜNTHER, A., MOL, F., GEROWITT, B., 2012, Analysing the influence of crop management on pesticide use intensity while controlling for external sources of variability with Linear Mixed Effects Models. *Agric. Syst.* 111:13–22;
- 5. CARMONA M., REIS E., CORTESE P., 1999, Leaf Spot of Wheat Diagnosis, Epidemiology And New Approache to The Management, *BASF*, *Buenos Aires*, Argentina;
- 6. COAKLEY, S. M., SCHERM, H., CHAKRABORTY, S., 1999, Climate change and plant diseases management. *Ann. Rev. Phytopathology*, 37:399-426;
- 7. COOLS H. J., FRAAIJE B. A., 2013, Update on mechanisms of azole resistance in *Mycosphaerella graminicola* and implications for future control. *Pest Manage. Sci.*69:150–155;
- 8. CORDO C. A., SIMÓN M. R., PERELLO A. E., ALIPPI H. E., 1999, Spore dispersal of leaf blotch of wheat *Mycosphaerella graminicola* and *Septoria tritici*, in *Septoria* and *Stagonospora* Diseases of Cereals. A Compilation of Global Research, M. van Ginkel, A. McNab, and J. Krupinsky, Eds., pp. 98–101, *CIMMYT*, *Ciudad de Mexico*, D.F., Mexico;
- 9. CORDO C. A., MÓNACO C. I., ALTAMIRO R., PERELLA A. E., LARRÁN S., KRIPELZ N. I., SIMÓN M. R., 2017, Weather conditions associated with the release and dspersal of *Zymoseptoria tritici* in the Argentine Pampas region. *International Journal of Agronomy*, vol. 217, p.13;
- 10. Dean R., Van Kan J. A. L., Pretorius Z. A., Hammond Kosack K. E., Di Pietro A., Spanu P. D., Rudd J. J., Dickman M., Kahmann R., Ellis J., Foster G. D., 2012, The top 10 fungal pathogens in molecular plant pathology. *Mol. Plant Pathol.* 13:414–430;
- 11. DHILLON B., GILL N., HAMELIN R. C., GOODWIN S. B., 2014, The landscape of transposable elements in the finished genome of the fungal wheat pathogen *Mycosphaerella graminicola*. *BMC Genomics*. 15:1132;
- 12. DUNCAN K. E., HOWARD R. J., 2000, Cytological analysis of wheat infection by the leaf blotch pathogen *Mycosphaerella graminicola*. *Mycol. Res.*104:1074–1082;
- 13. DUVIVIER M., DEDEURWAERDER G., DE PROFT M., MOREAU J. M., LEGREVE A., 2013, Real-time PCR quantification and spatio-temporal distribution of airborne inoculum of *Mycosphaerella graminicola* in Belgium. *European Journal of Plant Pathology* 137: 325–41;
- 14. EYAL, Z., AMIRIZ, WAHL, I., 1973, Physiologic specialisation of *Septoria tritici*. *Phytopathology* 63: 1087–1091;
- 15. EYAL Z., SCHAREN A. L., PRESCOTT J. M., VAN GINKEL M., 1987. The *Septoria* Diseases of Wheat, Concepts and Methods of Disease Management. *CIMMYT; Mexico*, D.F.;
- 16. Fernandez, M. R., Stevenson, C. F., Hodge, K., Dokken Bouchard, F., Pearse, P. G., Waelchli, F., Brown, A.; Peluola, C., 2016, Assessing effects of climatic change, region and agronomic practices on leaf spotting of bread and durum wheat in the Western Canadian Prairies, from 2001 to 2012. *Agron. J.* 108: 1180–1195;

- 17. GHINI, R., HAMADA, E., 2008, Mudanças climaticas: impactos sobre doenças de plantas no Brasilia: *Embrapa/SCT*, 331;
- 18. GHINI, R., HAMADA, E., BETTIOL, W., 2008, Climate Change and Plant Diseases. *Sci. Agric.* 65, special issue, 98-107;
- 19. GILBERT, J. AND WOODS, S. M., 2001, Leaf spot diseases of spring wheat in southern Manitoba farm fields under conventional and conservations tillage. *Can. J. Plant Sci.* 2001, 81, 551–559;
- 20. GLADDERS P., PAVELEY N. D., BARRIE I. A., HARDWICK N., HIMS M. J, LANGTON S., TAYLOR M. C., 2001, Agronomic and meteorological factors affecting the severity of leaf blotch caused by *Mycosphaerella graminicola* in commercial wheat crops in England. *Ann. Appl. Biol.*, 138: 301-311;
- 21. GOODWIN S. B., M' BAREK S. B., DHILLON B., WITTENBERG A. H., CRANE C. F., HANE J. K., 2011, Finished genome of the fungal wheat pathogen *Mycosphaerella graminicola* reveals dispensome structure, chromosome plasticity, and stealth pathogenesis. *PLoS Genet*.7:e1002070;
- 22. GOUACHE D., BENSDOUN A., BRUN F., OAGE C., MAKOWSKI D., WALLACH D., 2013, Modelling climate change impact on *Septoria tritici* Blotch (STB) in France: accounting for climate model and disease model uncertainty. *Agric. For. Meteorol.*, 170: 242-252;
- 23. GURR S. J., FONES H., 2015, The impact of *Septoria tritici* Blotch disease on wheat: an EU perspective. *Fungal Genet. Biol.*79:3–7;
- 24. HARVELL, C. D., MITCHELL, C. E., WARD, J. R., ALTIZER, S., DOBSON, A. P., OSTFELD, R. S., SAMUEL, M. D., 2002, Climate warming and disease risks for terrestrial and marine biota. *Science*, 296 (5576), 2158-2162. DOI: 10.1126/science.1063699;
- 25. JØRGENSEN L. N., HOVMØLLER M. S., HANSEN J. G., LASSEN P., CLARK B., BAYLES R., *et al.*, 2014, IPM Strategies and Their dilemmas including an introduction to www.eurowheat.org. *J. Integr. Agric.* 13: 265–281;
- 26. KETTLES, G. J., KANYUKA, K., 2016, Dissecting the molecular interactions between wheat and the fungal pathogen *Zymoseptoria tritici*. Front. Plant Sci. 7:508;
- 27. KELLNER R., BHATTACHARYYA A., POPPE S., HSU T. Y., BREM R. B., STUKENBROCK E. H., 2014, Expression profiling of the wheat pathogen *Zymoseptoria tritici* reveals genomic patterns of transcription and host-specific regulatory programs. *Genome Biol. Evol.* 6: 1353–1365;
- 28. KEMA G. H. J., YU D., RIJKENBERG F. H. J., SHAW M. W., BAAYEN R. P., 1996, Histology of the pathogenesis of *Mycosphaerella graminicola* in wheat. *Phytopathology*, 86:777–786;
- 29. Loarie, S. R., Duffy, P. B., Hamilton, H., Asner, G. P., Field G. B., Ackerly, D. D., 2009, The velocity of climate change. *Nature*, 462:1052-1055;
- 30. LOVELL D. J., PARKER S. R., HUNTER T., ROYLE D. J., AND COKER R. R., 1997, Influence of crop growth and structure on the risk of epidemics by *Mycosphaerella graminicola (Septoria tritici)* in winter wheat, *Plant Pathology*, vol. 46, no. 1:126–138;
- 31. LOVELL D. J., HUNTER T., POWERS S. J., PARKER S. R., VAN DEN BOSCH F., 2004, Effect of temperature on latent period of *Septoria* leaf blotch on winter wheat under outdoor conditions. *Plant Pathology* 53: 170–181;
- 32. McDonald B. A., Zhan J., Yarden O., Hogan K., Garton J., Pettway R. E., 1999, The population genetics of *Mycosphaerella graminicola* and *Stagonospora nodorum*. In: Lucas J.A., Bowyer P., Anderson H.M., editors. *Septoria* on Cereals: A Study of Pathosystems. *CABI Publishing*; Wallingford, UK, pp. 44–69;
- 33. MEHRABI R., VAN DER LEE T., WAALWIJK C., KEMA G. H. J., 2006, MgSlt2, a cellular integrity MAP kinase gene of the fungal wheat pathogen *Mycosphaerella graminicola*, is dispensable for penetration but essential for invasive growth. *Mol. Plant Microbe Interact*. 19:389–398;
- 34. MEHRABI R., ZWIERS L. H., DE WAARD M. A., KEMA G. H., 2009, MgHog1 regulates dimorphism and pathogenicity in the fungal wheat pathogen *Mycosphaerella graminicola*. *Mol. Plant Microbe Interact*.19:1262–1269;
- 35. MIRZADI GOHARI A., MEHRABI R., ROBERT O., INCE I. A., BOEREN S., SCHUSTER M., STEINBERG G., DE WIT P. J., KEMA G. H., 2014, Molecular characterization and functional analyses of ZtWor1, a

- transcriptional regulator of the fungal wheat pathogen *Zymoseptoria tritici*. *Mol. Plant Pathol*.15:394–405:
- 36. Mundt C. C., Brophy L. S., Schmitt, 1999, Choosing crop cultivars and cultivar mixtures under low versus high disease pressure: a case study with wheat, *Crop Protection*, 14:509-515;
- 37. MUNDT C. C., COWGER, GARRETT K. A., 2002, Relevance of integrated disease management to resistance durability, *Euphytica*, 124: 245-252;
- 38. NEWTON A. C., JOHNSON S. N., GREGORY P. J., 2011, Implications of climate change for diseases, crop yields and food security. *Euphytica* 179:3-18, NOAA report, Jan. 2017;
- 39. ORTON E. S., DELLER S., BROWN J. K., 2011, *Mycosphaerella graminicola*, from genomics to disease control. *Mol. Plant Pathol.*, 12:413–424;
- 40. PIETRAVALLE S., SHAW M. W., PARKER S. R., VAN DEN BOSCH F., 2003, Modeling of relationships between weather and *Septoria tritici* epidemics on winter wheat: a critical approach. *Phytopathology*, 93: 1329-1339;
- 41. PONOMARENKO, A., GOODWIN, S. B., KEMA., G. H. J., 2011, *Septoria Tritici* Blotch (STB) of Wheat. Available online at: https://www.apsnet.org/edcenter/intfungi/ascomycetes/Pages/Septoria.aspx [Accessed on 15th January, 2018];
- 42. POPP, J., PETÖ, K., NAGY, J., 2013, Pesticide productivity and food security. A review. Agronomy for Sustainable Development, vol. 33 (1): 243-255;
- 43. PRESTES, A. M., HENDRIX, J. W., 1978, The role of *Stellaria media* in the epidemiology of *Septoria tritici* on wheat. In: *3rd International Congress of Plant Pathology ISPP*, p. 336;
- 44. RUDD J. R., KANUKA K., HASSANI-PAK K., DERBYSHIRE M., ANDONGABO A., DEVONSHIRE J., LYSENKO A., SAQU M., DESAI N. M., POWERS S. J., HOOPER J., AMBROSO L., BHARTI A., FARMER A., HAMMOND KOSACK K. E., DIETRICH R. A., COURBOT M., 2015, Transcriptome and metabolite profiling the infection cycle of *Zymoseptoria tritici* on wheat reveals a biphasic interaction with plant immunity involving differential pathogen chromosomal contributions and a variation in the hemibiotrophic lifestyle definition. *Plant Physiol*. doi: 10/pp114.255927;
- 45. SCHUH W., 1990, Influence of tillage systems on disease intensity and spatial pattern of *Septoria* leaf blotch, *Phytopathology*, vol. 80:1337–1340;
- 46. SIMÓN M. R., PERELLÓ A. E., CORDO C. A. et al., 2005 Association between Septoria tritici blotch, plant height and heading date in wheat, Agronomy Journal, vol. 97, no. 4: 1072–1081;
- 47. SAINTENAC, C., WING SHAM, L., CAMBON, F., RUDD, J. J., KING, R. C., MARANDE, W., POWERS, S. S., BERGÈS, H., PHILLIPS, A. L., MANY, C., KOSACK, K. E. H., LONGIN T., KANYUKA, K., 2018, Wheat receptor kinase -like protein Stb6 controls gene-for-gene resistance to fungal pathogen *Zymoseptoria tritici*. *Nature Genetics*, 50:368-374;
- 48. STEINBERG, G., 2015, Cell biology of *Zymoseptoria tritic*i> pathogen cell organization and wheat infection. *Fungal Genet. Biol.* 79:17-23;
- 49. STUKENBROCK E. H., BATAILLON T., DUTHEIL J. Y., HANSEN T. T., LI R., ZALA M., *et al.*, 2011, The making of a new pathogen: insights from comparative population genomics of the domesticated wheat pathogen *Mycosphaerella graminicola* and its wild sister species. *Genome Res.* 21: 2157–2166;
- 50. STUKENBROCK E. H., JORGENSEN F. G., ZALA M., HANSEN T. T., MCDONALD B. A., SCHIERUP M. H., 2006, Whole-genome and chromosome evolution associated with host adaptation and speciation of the wheat pathogen *Mycosphaerella graminicola*. *PloS Genet*. 6:e1001189;
- 51. SUFFERT F., SACHE I., LANNOU C., 2011, Early stages of *Septoria tritici* blotch epidemics of winter wheat, build-up, overseasoning, and release of primary inoculum. *Plant Pathol.*, 60:166–177;
- 52. SUFFERT F., DELESTRE G., GÉLISSE L., 2018, Sexual reproduction in the fungal foliar pathogen *Zymoseptoria tritici* is driven by antagonistic density-dependence mechanisms. *Microbial Ecology on line* at https://doi.org/10.1101/290072 [Accessed on 26th January, 2018];
- 53. TE BEEST D. E., SHAW M. W., PAVELEY N. D., VAN DEN BOSCH F., 2009, Evaluation of a predictive model for *Mycosphaerella graminicola* for economic and environmental benefits. *Plant. Pathol.*, 58:1001-1009;

- 54. TORRIANI S. F. F., BRUNNER P. C., MCDONALD B. A., SIEROTZI S., 2009, Q01 resistance merged independently at least four times in European populations of *Mycosphaerella graminicola* Pest *Manag. Sci.*, 65:155-162;
- 55. TORRIANI, S. F., MELICHAR, J. P., MILLS, C., PAIN, N., SIEROTZKI, H., COURBOT, M., 2015, *Zymoseptoria tritici:* a major threat to wheat production, integrated approaches to control. *Fungal Genet. Biol.* 79: 8–12;
- 56. VÁRY, Z., MULLINS, E., MCELWAIN, J. C. AND DOOHAN, F. M., 2015, The severity of wheat diseases increases when plants and pathogens are acclimatized to elevated carbon dioxide. *Global Change Biology*, 21: 2661–2669;
- 57. WALTERS, D. R. AND BINGHMANN, I. J., 2007, Influence of nitrogen on disease development caused by fungal pathogens: Implications for plant disease control. J. Appl. Biol. 151:307-324;
- 58. Wenda Piesik, A., Lemańczyk, G., Pańka, D., Piesik, D., 2016, Risk assessment posed by diseases in context of integrated management of wheat. *J. Plant Dis. Prot.* 123: 3–18;
- 59. WITTENBERG A. H., VAN DER LEE T. A., BEN M'BAREK S., WARE S. B., GOODWIN S. B., KILIAN A., VISSER R. G., KEMA G. H., SCHOUTEN H. J., 2009, Meiosis drives extraordinary genome plasticity in the haploid fungal plant pathogen *Mycosphaerella graminicola*. *PloS One*. 4:e5863;